Pearls of Glaucoma Management

JoAnn A. Giaconi Simon K. Law Kouros Nouri-Mahdavi Anne L. Coleman Joseph Caprioli *Editors*

Second Edition



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ISBN 978-3-662-49040-2 ISBN 978-3-662-49042-6 (eBook) DOI 10.1007/978-3-662-49042-6

Library of Congress Control Number: 2016941933

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Printed on acid-free paper

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Foreword

If you have ever uttered the commonly expressed lament, "Glaucoma is so confusing!" then this text is for you. You will no longer be bewildered.

Why practitioners may be confused about how to be of help to patients with glaucoma—in its many incarnations and reincarnations—is easily understood. The issue seems to be overwhelming when one considers that the already massive population of those with glaucoma is increasing rapidly as the world's population increases and ages.

During the past 50 years the fundamental definition of glaucoma has changed almost 180°, and the indications for treatment have become more variable and controversial, some advising early therapy and others strongly cautioning against such an approach: Various diagnostic tests have come and gone and are interpreted in such different ways that there seems to be no consensus; surgical techniques come in and out of fashion in perplexing ways. There seems to be a constantly shifting, sandy foundation on which are built unsteady schools of ever-varying advice. Why practitioners, patients, and the public are often bewildered is understandable.

The current text was designed to be relevant, scientific, and practical. The editors have accomplished their objective well. The authors chosen to share their wisdom are expert practitioners who recognize the dangers of basing treatment on theory. They, the leaders in their fields, create an understanding of glaucoma and conditions related to glaucoma that is sound, scientific, and effective. The editors clearly instructed their contributors to avoid speculation, to be practical, and to insist on evidence, not opinion (and where good evidence was lacking, to indicate such a lack). The result is a cohesive picture that should be of immense help to all those trying to make sense of what to many seems to be confusing.

It is perhaps not surprising that this text accomplishes its objective so admirably. The senior editor is a vastly experienced physician, equally at home in the clinic, the operating room, the classroom, and in a basic research laboratory. The contributing authors come from many different institutions and cultures; some are younger and others older. The current text, however, does not present information that must be sifted by a discerning reader in order to come up with appropriate advice. Rather, the authors simplify, clarify, organize, and explain practically and scientifically. Those wanting to know how to approach patients with glaucoma or those many, many patients in whom it is not clear whether glaucoma is present or not will find this a treasure trove of sound science blended with critical experience. The need for this intellectually vigorous, practical approach to caring for patients with conditions related to intraocular pressure and optic nerve disease is great. There is probably truth in the belief that all persons will eventually develop glaucoma if they live long enough. As the world population ages and increases, as resources become ever more precious, and as cost considerations become more confining, there is increasing urgency for guidelines that concentrate on the essentials and that will help achieve the goal of caring for the sick and for the well, specifically, the greatest good for the greatest number, while still addressing the needs and wants of each individual person.

Currently there is much interest in "translational research." This book is highly successful in translating vast amounts of disparate, sometimes disconcerting information into understandable sentences, paragraphs, and illustrations that will result in more effective and more relevant care.

Philadelphia, PA, USA

George Spaeth

Preface

This book was developed based on the questions that clinicians, fellows, and residents taking care of glaucoma patients have asked us as consultants. Most textbooks on glaucoma provide a broad overview of the clinical and basic science literature, which is very useful to students learning about glaucoma. However, these textbooks may leave many questions unanswered for the clinician searching for advice on how to manage a specific problem. This book asks and answers those questions. Additionally, it covers topics that are not always included in traditional textbooks but that are being discussed at national and international meetings.

In addition to asking the questions that frequently arise in managing patients with glaucoma, a goal of this textbook was to have the authors who are familiar with the world literature digest that information in the context of their own clinical experience. We asked authors to answer questions the way they might answer a physician's questions over the phone. We asked them to state their opinions on how they like to manage clinical situations, where appropriate, and to also point out that their preferred management is not the only way to manage the problem if other acceptable means are available. The questions are organized by topic and cover diagnostic testing and interpretation, risk factors, medical treatment, procedural treatments, various glaucoma subtypes, and complications.

We must thank all the consulting physicians, students, residents, and fellows who we have encountered and who inspired this textbook. As well, we thank Ms. Minn Oh for administrative help with the second edition of this book.

Los Angeles, CA, USA JoAnn A. Giaconi Simon K. Law Anne L. Coleman Kouros Nouri-Mahdavi Joseph Caprioli

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Optic Nerve: The Glaucomatous Optic Nerve

Claude F. Burgoyne

Core Messages

- The principle insult in glaucoma occurs within the neural, cellular, and connective tissues of the optic nerve head (ONH).
- Intraocular pressure at all levels has biomechanical effects on the optic nerve tissues.
- Clinical cupping is one manifestation of the pathophysiology of glaucomatous damage, but is not the pathophysiology itself.
- The variable appearance of the ONH in all optic neuropathies is the predictable result of ONH tissue biomechanics.
- As our clinical tools for characterizing ONH biomechanics improve, so too will our ability to understand normal ONH aging and its contributions to the clinical behavior and susceptibility of the ONH.

C.F. Burgoyne, M.D. (🖂)

1.1 Why Is the Optic Nerve Important in the Diagnosis and Management of Glaucoma?

Glaucoma is an optic neuropathy. Although there are several pathophysiologies that must be managed in the clinical care of the glaucoma patient, what defines all forms of glaucoma is an optic neuropathy that demonstrates classic and recognizably variable [1–6] structural and functional behaviors.

1.1.1 The Optic Nerve Head Is the Principal Site of Glaucomatous Damage to the Visual System

Although glaucomatous damage likely encompasses important pathophysiology within the retinal ganglion cell (RGC) stroma [7–12], photoreceptors [13–17], lateral geniculate body [18–20], and visual cortex [20], strong evidence suggests that damage to the RGC axons within the lamina cribrosa of the optic nerve head (ONH) [21–26] is the central pathophysiology underlying glaucomatous vision loss. Recent studies in monkeys [25–30], rats [31–33], and mice [34] support the importance of the ONH in glaucoma by describing profound alterations at

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the earliest detectable stage of the disease within the prelaminar, laminar, and peripapillary scleral tissues of the ONH.

The ONH tissues make up a dynamic environment wherein 1.2-2.0 million RGC axons converge, turn, and exit the eye through the inner (Bruch's membrane opening) and outer (scleral) portions of the neural canal (Fig. 1.1). Within the scleral portion of the canal, the bundled axons pass through a three-dimensional meshwork of astrocyte-covered, capillary-containing connective tissue beams known as the lamina cribrosa (Fig. 1.1). Within the lamina, axonal nutrition is dependant upon the movement of oxygen and nutrients from the laminar capillaries, through the laminar beam extracellular matrix (ECM), into the laminar astrocyte processes within the beam, finally reaching the peripheral and central axons of each bundle, via cell processes [35].

The connective tissue beams of the lamina cribrosa are anchored via the neural canal wall to a circumferential ring of collagen and elastin fibers within the peripapillary sclera [36–38] and are presumed to bear the forces generated by

intraocular pressure (IOP) (Fig. 1.1). IOP-related stress (force/cross-sectional area of the tissue experiencing that force) and strain (a measure of local deformation of a tissue induced by applied stress) within the load-bearing tissues of the ONH influence the physiology and pathophysiology of all three ONH tissue types (Table 1.1): (1) the connective tissues, (2) the neural tissues, and (3) the cells that exist alone or in contact with both (1) and (2) [39–41].

While the pathophysiology of glaucomatous damage to the ONH tissues remains controversial, we have proposed that it is multifactorial and is influenced by at least three etiologies (Table 1.2)—IOP-related connective tissue stress and strain [21–24], blood flow/nutrient diffusion/ ischemia within the laminar and prelaminar tissues [42–45], and the autoimmune and/or inflammatory state of the tissues [46–51] (Fig. 1.2, top). The interplay between the pathophysiology of ONH neural and connective tissue damage and the clinical appearance and behavior of the neuropathy are discussed in Figs. 1.2 and 1.3 and the sections that follow.

Fig. 1.1 The optic nerve head (ONH) is centrally influenced by IOP-related stress and strain. The ONH is made up of prelaminar, laminar, and retrolaminar regions (a). Within the clinically visible surface of the normal ONH (referred to as the optic disc) (b), central retinal vessels enter the eye and retinal ganglion cell (RGC) axons appear pink because of their capillaries (which are principally supplied by branches from the posterior ciliary arteries (PCA) in (c). The primary site of RGC axon insult in Glaucoma is within the lamina cribrosa (schematically depicted with axon bundles) in (d), isolated by trypsin digest in a scanning electron micrograph in (e) and drawn with stippled extracellular matrix (ECM), central capillary (red), and surrounding astrocytes (yellow with basement membranes in black) (f). Blood flow within the ONH, while controlled by autoregulation, can be affected by non-IOP-related effects such as systemic blood pressure fluctuation and vasospasm within the retrobulbar portion of the PCAs. Additional IOP-induced effects may include compression of PCA branches within the peripapillary sclera (due to scleral stress and strain) and compression of laminar beam capillaries reducing laminar capillary volume flow (c, f) [43]. There is no direct blood supply to the axons within the laminar region. Axonal nutrition within the lamina (f) requires diffusion of nutrients from the laminar capillaries, across the endothelial and pericyte basement membranes, through the ECM of the laminar beam, into astrocyte processes within the beam, through the astrocyte processes into the adjacent axons (vertical lines). Chronic age-related changes in the endothelial cell and astrocyte basement membranes, as well as IOP-induced changes in the laminar ECM and astrocyte basement membranes may diminish nutrient diffusion to the axons in the presence of a stable level of laminar capillary volume flow. The clinical manifestation of IOP-induced damage to the ONH is most commonly "deep cupping" (g), but in some eyes cupping can be shallower accompanied by pallor (h). Z-H circle of Zinn-Haller; PCA posterior ciliary arteries; NFL nerve fiber layer; PLC prelaminar region; LC lamina cribrosa; RLC retrolaminar region; ON optic nerve; CRA central retinal artery. (a) Reproduced with permission of Arch Ophthalmol. Copyright 1969 American Medical Association. All Rights reserved [35]. (b, g, h) Reprinted with permission from J Glaucoma. Copyright 2008 [83]. (c) Reprinted with permission from Elsevier. Copyright 1996. This article was published in The Glaucomas. Edited by Ritch R, Shields MB, Krupin T. Mosby, St. Louis; Cioffi GA, Van Buskirk EM: Vasculature of the anterior optic nerve and peripapillary choroid. Pg 177-197 [140]. (d) Courtesy of Harry A. Quigley and reprinted with permission from Kugler Publications, Amsterdam [141]. (e) Reproduced with permission of Arch Ophthalmol. Copyright 1990 American Medical Association. All Rights reserved. (f) Reproduced with permission of Arch Ophthalmol. Copyright 1989 American Medical Association. All Rights reserved [142]



Table 1.1 Optic nerve head tissue types

- Connective tissues

 Load-bearing connective tissues of the peripapillary sclera, scleral canal wall, and lamina cribrosa

 Neural tissues

 Retinal ganglion cell (RGC) axons
 Cells that exist alone or in contact with 1 and 2 above
 Astrocytes
 Glial cells
 Endothelial cells
- Pericytes
- Basement membranes (BM)

 Table 1.2 Primary proposed etiologies glaucomatous damage to the ONH

IOP-related connective tissue stress and strain

Blood flow/nutrient diffusion and/or ischemia within the laminar and prelaminar tissues

Autoimmune and/or inflammatory mechanisms within the tissue

Retro Etiology Auto-immune hulbar Blood flow and **IOP-Related** IOP terminants or Nutrient supply stress/strain Inflammatory of (CSF) Within the ONH Insults ONH blood flow Axonal **Astrocytes/Glia** ONH damage connective Within the Tissue lamina damage Cribrosa by multiple by multiple Pathophysiology mechanisms mechanisms Prelaminar 1° or 2° Neural **IOP-Induced** CT Tissue deformation thinning Prelaminar Laminar component of component of cupping cupping Stiffness Variable ONH CT More compliant Less compliant 5 Appearance of the Stiffness (Old) (Young) **ONH** in glaucoma (Age)

Fig. 1.2 While damage to the neural and connective tissues of the ONH is multifactorial, ONH appearance in the neuropathy is importantly influenced by connective tissue stiffness. In our biomechanical paradigm, IOP-related strain influences the ONH connective tissues and the volume flow of blood (primarily) and the delivery of nutrients (secondarily), through chronic alterations in connective tissue stiffness and diffusion properties (explained in Fig. 1.1). Non-IOP-related effects such as autoimmune or inflammatory insults (*yellow*) and retrobulbar determi-

nants of ocular blood flow (*red*) can primarily damage the ONH connective tissues and/or axons, leaving them vulnerable to secondary damage by IOP-related mechanisms at normal or elevated levels of IOP. Once damaged, the ONH connective tissues can become more or less rigid depending upon lamina cribrosa astrocyte and glial response. If weakened, ONH connective tissues deform in a predictable manner, which underlies a laminar component of clinical cupping (Figs. 1.3 and 1.4). Reprinted with permission from J Glaucoma, copyright 2008 [83]



Fig. 1.3 All clinical cupping, regardless of etiology, is a manifestation of underlying "prelaminar" and "laminar" pathophysiologic components. (a) Normal ONH. To understand the two pathophysiologic components of clinical cupping, start with (b) a representative digital central horizontal section image from a postmortem 3D reconstruction of this same eye (white section line in (a))-vitreous top, orbital optic nerve bottom, lamina cribrosa between the sclera and internal limiting membrane (ILM) delineated with green dots. (c) The same section is delineated into principle surfaces and volumes (black-ILM; purple-prelaminar neural and vascular tissue; cyan blue line-bruchs membrane opening (BMO)-zero reference plane cut in section; green outline-post-BMO total prelaminar area or a measure of the space below BMO and the anterior laminar surface). (d) Regardless of the etiology, clinical cupping can be "shallow" (e) or "deep" (f) (these clinical photos are representative and are not of the eye in (a)). A prelaminar or "shallow" form of cupping (g, black arrows) is primarily

due to loss (thinning) of prelaminar neural tissues without important laminar or ONH connective tissue involvement. Laminar or "deep" cupping (h, small white arrows depict expansion of the green shaded space) follows ONH connective tissue damage and deformation that manifests as expansion of the total area beneath BMO, but above the lamina. Notice in (h) that while a laminar component of cupping predominates (white arrows) there is a prelaminar component as well (black arrows). While prelaminar thinning is a manifestation of neural tissue damage alone, we propose that laminar deformation can only occur in the setting of ONH connective tissue damage followed by permanent (fixed) IOP-induced deformation (Reprinted with permission from [30]). Investigative Ophthalmology & Visual Science by Hongli Yang. Copyright 2007 by Investigative Ophthalmology & Visual Science. Reproduced with permission of Investigative Ophthalmology & Visual Science in the format Textbook via Copyright Clearance Center [30]



Fig. 1.4 Our central hypothesis regarding ONH connective tissue damage in "laminar" cupping. "Deep," "laminar," or "glaucomatous" cupping is a manifestation of ONH connective tissue damage, which can be caused by either IOP-related or non-IOP-related insults (see Fig. 1.5). However, regardless of the primary insult to the ONH connective tissues, their deformation (if present) is driven by IOP-related connective tissue stress and strain. Thus, the presence of ONH connective tissue deformation in any optic neuropathy is evidence that the level of IOP at which it occurred (whether normal or elevated) is too high for the connective tissues in their present condition. (a) Schematic of normal laminar thickness (x) within the scleral canal with scleral tensile forces acting on the scleral canal wall. (b) Early IOP-related damage in the monkey eye [25–30] includes posterior bowing of the lamina and peripapillary sclera accompanied by neural canal expansion (mostly within the posterior (outer) scleral portion) and thickening

(not thinning) of the lamina (y). In our studies to date, this appears to represent mechanical yield (permanent stretching) rather than mechanical failure (physical disruption) of the laminar beams (c). Progression to end-stage damage includes profound scleral canal wall expansion (clinical excavation) and posterior deformation and thinning of the lamina (z) by mechanisms that are as yet uncharacterized [143, 144]. If all other aspects of the neuropathy are identical, the stiffer the lamina, the more resistant it will be to deformation. Whether this is better or worse for the adjacent axons is a separate question that remains to be determined. Reprinted from Prog Retin Eye Res:24. Burgoyne CF, Downs JC, Bellezza AJ, Suh JK, Hart RT: The optic nerve head as a biomechanical structure: a new paradigm for understanding the role of IOP-related stress and strain in the pathophysiology of glaucomatous optic nerve head damage; pp. 39-73. Copyright (2005) with permission from Elsevier [41]

1.1.2 The Pathophysiology of Glaucomatous Damage Is Separate from the Clinical Phenomenon of "Cupping"

Cupping is a clinical term used to describe enlargement of the ONH cup in all forms of optic neuropathy [52–59]. However, *cupping* is also used as a synonym for the pathophysiology of glaucomatous damage to the ONH [24, 60–62]. Because the clinical and pathophysiologic contexts for *cupping* are seldom clarified, there is a confusing literature regarding the presence, importance, and meaning of *cupping* in a variety of optic neuropathies [2, 63–76].

We have previously proposed [30] that all optic neuropathies can demonstrate clinical cupping and that all forms of *clinical* cupping have two principal pathophysiologic componentsprelaminar thinning and laminar deformation (Fig. 1.3). Prelaminar thinning results from net thinning of the prelaminar tissues due to physical compression and/or loss of RGC axons even in the presence of gliosis [77-80]. In this paradigm, prelaminar thinning results in a clinically shallow form of cupping [81, 82] (being limited to the prelaminar tissues) that occurs in all forms of RGC axon loss (including aging) and is therefore nonspecific. Laminar deformation results in a clinically deeper form of cupping that occurs only in those optic neuropathies in which damaged ONH connective tissues (lamina cribrosa and peripapillary scleral connective tissue) have become susceptible to permanent, IOP-induced deformation [25, 26, 28, 29, 41]. Whether the ONH connective tissues are primarily damaged by IOP or some other insult (ischemic, autoimmune, inflammatory, secondary astrocyte activation, or genetic predisposition [41]) (Fig. 1.4), if they deform they do so under the effects of IOP (normal or elevated) in a predictable way, and this deformation underlies laminar or deep or glaucomatous cupping (Figs. 1.3 and 1.4).

The previous paragraph contains two important ideas. First, it is possible for non-IOP-related processes to damage the ONH primarily and still end up with a nerve that looks and behaves in a manner we call *glaucomatous*. Second, IOPrelated connective tissue stress and strain still drive the processes that cause the damaged tissues to deform, even if IOP is not the primary insult in the process and regardless of whether IOP is high or low.

1.1.3 The Clinical Appearance and Behavior of the ONH Holds Clues as to the Etiology of a Given Optic Neuropathy

When IOP is not elevated, and sometimes even when it is, the clinical challenge in the examination of the optic disc is not to recognize glaucoma, but rather to recognize the presence of an optic neuropathy and then separately determine the likelihood that IOP is playing a contributing role. The notions of laminar and prelaminar cupping suggest two important concepts to consider in the clinical assessment of an optic neuropathy.

First, detection of clinical cupping or its progression suggests the presence of an optic neuropathy, but it does not confirm that IOP is the etiologic agent. Regardless of clinical circumstances, but particularly when IOP is within normal limits, clinical cupping without clinically detectable connective tissue deformation should not be an absolute indication for IOP lowering. We have previously proposed that in patients with robust ONH connective tissues. IOP-related stress and strain can cause a prelaminar form of cupping in which pallor exceeds excavation by causing axonal degeneration without damage to the underlying connective tissues [41, 83]. Having proposed this concept, we now emphasize that without direct evidence of ONH connective tissue damage, the role of IOP in an individual optic neuropathy cannot be certain.

Second, in contrast to surface change detection, clinical detection of ONH connective tissue damage (i.e., a "laminar" contribution to cupping) is direct evidence of IOP involvement in the neuropathy and should become an absolute indication for IOP lowering, regardless of the level of IOP or the etiology of the primary connective tissue insult (ischemia, autoimmune, inflammatory, or IOP-related strain) [41, 83]. Thus, in all eyes, the presence of laminar cupping has diagnostic significance if we can develop the clinical tools to detect it.

1.1.4 The Aged ONH Holds Important Clues About Susceptibility

A variety of data suggest that the ONH becomes more susceptible to progressive glaucomatous damage as it ages, though this concept remains unproven through direct experimentation and it may not hold true for every aged eye. The data to date can be summarized as follows. First, in most [84-88] but not all [89, 90] population-based studies, IOP does not increase with age, and in some studies where it does increase, the magnitude of increase is not likely to be clinically important. Thus, the fact that the prevalence of glaucoma increases with age [91–93] is likely explained by a greater susceptibility to IOP and other non-IOP-related risk factors, rather than to a higher prevalence of IOP elevation with increasing age. Second, in an extensive review of the literature, low-tension glaucoma is a disease of the elderly [94–99], with only a few reports regarding the onset and progression of normal tension glaucoma in infants, children, and young adults [100]. Third, age is an independent risk factor for both the prevalence [91-93] and progression of the neuropathy at all stages of damage [101–103].

1.1.5 How Age Influences the Susceptibility and Clinical Behavior of the ONH

Over a lifetime, the ONH connective tissues are exposed to substantial levels of IOP-related stress and strain at normal levels of IOP. This stress and strain increases as IOP increases and/or fluctuates (Fig. 1.5) [104–108]. Stresses and strains at a given level of IOP are physiologic or pathophysiologic depending upon the response of the tissues that experience them (Fig. 1.5). In this context, IOP is not so much normal as physiologic or pathophysiologic and what constitutes physiologic and pathophysiologic levels for IOP may change as they are influenced by associated systemic factors and aging.

Physiologic stress and strain induce a broad spectrum of changes in both the connective



Fig. 1.5 Over the course of a lifetime, whether an eye demonstrates the "neuropathy of aging" or the neuropathy of glaucoma lies in ONH susceptibility. For a given ONH, IOP generates low or high levels of stress depending upon the 3D architecture of the ONH connective tissues (size and shape of the canal, thickness of the lamina and sclera—*susceptibility 1*). Some ONHs will have relatively low stress at high IOP (*d*). Others will have high stress at low IOP (*e*). Whether a given level of IOP-related stress is physiologic or pathophysiologic depends upon the ONH's microenvironment (*susceptibility 2*). Strong connective tissues, a robust blood supply, and stable astrocytes and

glia increase the chance of normal ONH aging (*right, bot-tom*). While the existence of a neuropathy of aging is controversial, the difference between "normal" age-related axon loss (if it is shown to exist) and the development of glaucomatous damage is a matter of ONH susceptibility (Reprinted with permission from [41]). Reprinted from Prog Retin Eye Res:24. Burgoyne CF, Downs JC, Bellezza AJ, Suh JK, Hart RT: The optic nerve head as a biomechanical structure: a new paradigm for understanding the role of IOP-related stress and strain in the pathophysiology of glaucomatous optic nerve head damage; pp. 39–73. Copyright (2005) with permission from Elsevier [41]

tissues and vasculature that are central to normal aging. While the concepts of age-related optic nerve axon loss [33, 109–114] and an optic neuropathy of aging [2, 55, 113–115] remain controversial, we believe that the range of physiologic stress and strain experienced within the ONH connective tissues over a lifetime are likely to be of central importance to both concepts.

Pathophysiologic stress and strain induce pathologic changes in cell synthesis and tissue microarchitecture (Fig. 1.5) that exceed the effects of aging. These changes underlie two governing pathophysiologies in glaucoma: (1) mechanical yield and/or failure of the loadbearing ONH connective tissues (Figs. 1.2, 1.3, and 1.4), and (2) progressive damage to the adjacent axons by a variety of mechanisms (Fig. 1.2).

The aged ONH is more likely to have stiff connective tissues [116–128] and a compromised blood supply [129, 130]. However, age-related increases in laminar beam thickness [117, 120, 122, 127, 131], laminar astrocyte basement membrane thickness [120, 131], and laminar ECM hardening [117, 120, 122, 131] should not only increase laminar beam stiffness, but should also diminish nutrient diffusion from the laminar capillaries into adjacent axons (Fig. 1.1). Thus, for a given magnitude of IOP insult, the aged ONH should demonstrate (1) less deformation due to the presence of a stiffer lamina and peripapillary sclera and (2) more pallor for a given amount of deformation because (a) the aged ONH may be more susceptible to axon loss and (b) pallor precedes deformation in the aged eye, while deformation precedes (or supersedes) pallor in the young eye.

Apart from the issue of ONH susceptibility, we predict that if all aspects of insult are equal (alterations in IOP, the volume flow of blood and nutrient transfer from the laminar capillary to the ONH astrocyte are all of the same magnitude, duration, and fluctuation), the aged eye will demonstrate clinical cupping that is on average shallow and pale (at all stages of field loss) compared with the eye of a child or a young adult. This clinical behavior in its most recognizable form is described as *senile sclerotic cupping* [1–6, 132].

We thus propose an overlap between the optic neuropathy of aging and the optic neuropathy of glaucoma in the aged eye and a biomechanical explanation for why the aged eye should demonstrate a shallow form of clinical cupping in which pallor more than deformation predominates.

1.1.6 Apart from the Aged ONH, Are There Some Nerves That Are Mechanically More Sensitive to Damage?

Although IOP [133–136] has been shown to play a causative role in glaucomatous ONH damage at all levels of IOP, many questions remain. There is no agreement on the effects of IOP within the tissues of the ONH; no data exist that would allow one to predict a safe level of IOP for a given ONH; and there are no accepted explanations for the varied clinical manifestations of glaucomatous damage [3], glaucomatous cupping, and glaucomatous visual field loss.

The principal ocular determinants of ONH susceptibility to a given level of IOP are likely to include (1) the IOP level (both the magnitude and variation); (2) the geometry and material properties of the ONH and peripapillary scleral connective tissues; (3) the volume flow and perfusion pressure of blood within the laminar capillaries; (4) nutrient diffusion to the astrocytes for a given level of blood volume and pressure; (5) the molecular response of astrocytes and glia to physical strain within their basement membrane and the presence of physiologic stress within their microenvironment (Fig. 1.2); (6) RGC factors that make its axon more susceptible to damage within the ONH, or its stroma more susceptible to apoptosis in response to axonal distress; (7) the immune environment of the ONH and retina; and (8) the number of remaining viable axons.

At present, we lack the means to directly assess any of the determinants listed above; however, the following features may soon be within the reach of a variety of new imaging strategies and may contribute to clinically derived engineering finite element models of individual ONHs that we hope will one day underlie target pressure assignment: (1) the three-dimensional geometry and material properties of the lamina cribrosa, scleral flange, and peripapillary sclera [104–108]; (2) the difference in material properties between the peripapillary sclera and the lamina cribrosa [137, 138]; (3) the flow of blood and transport of nutrients across the basement membranes and ECM of the laminar beams; (4) the volume flow of blood through the intrascleral branches of the posterior ciliary arteries; and (5) the presence of peripapillary scleral posterior bowing and the distance between the anteriormost point of the subarachnoid space and the vitreous cavity [139].

Summary for the Clinician

- Glaucoma is an optic neuropathy in which the principal insult to the visual system is multifactorial and occurs within the neural, cellular, and connective tissues of the optic nerve head (ONH).
- IOP is a contributing risk factor to this pathophysiology at low, normal, and elevated levels because of its primary and secondary biomechanical effects on these tissues.
- Clinical cupping is one manifestation of the pathophysiology of glaucomatous damage, but is not the pathophysiology itself.
- A shallow form of cupping is nonspecific and can be expected to occur in all forms of optic neuropathy. Although the clinical appearance and behavior of the neuropathy of glaucoma can vary and include shallow forms of cupping, the pathophysiology of glaucomatous damage classically involves a deep form of cupping, which is a manifestation of ONH connective tissue damage and deformation. The variable appearance of the ONH in all optic neuropathies is the predictable result of ONH tissue biomechanics.

• As our clinical tools for characterizing ONH biomechanics improve, so too will our ability to understand normal ONH aging and its contributions to the clinical behavior and susceptibility of the ONH.

Acknowledgements Portions of this chapter and its figures appeared in the following article and are used with the permission of the *Journal of Glaucoma*:

Burgoyne CF, Downs JC. Premise and Prediction— How Optic Nerve Head Biomechanics Underlies the Susceptibility and Clinical Behavior of the Aged Optic Nerve Head. Invited original article. J Glaucoma 2008;17:318–328.

References

- Broadway DC, Nicolela MT, Drance SM. Optic disk appearances in primary open-angle glaucoma. Surv Ophthalmol. 1999;43 Suppl 1:S223–43.
- Jonas JB, Grundler A. Optic disc morphology in "age-related atrophic glaucoma". Graefes Arch Clin Exp Ophthalmol. 1996;234:744–9.
- Nicolela MT, Drance SM. Various glaucomatous optic nerve appearances: clinical correlations. Ophthalmology. 1996;103:640–9.
- Nicolela MT, Drance SM, Broadway DC, et al. Agreement among clinicians in the recognition of patterns of optic disk damage in glaucoma. Am J Ophthalmol. 2001;132:836–44.
- Nicolela MT, McCormick TA, Drance SM, et al. Visual field and optic disc progression in patients with different types of optic disc damage: a longitudinal prospective study. Ophthalmology. 2003;110: 2178–84.
- Nicolela MT, Walman BE, Buckley AR, Drance SM. Various glaucomatous optic nerve appearances. A color Doppler imaging study of retrobulbar circulation. Ophthalmology. 1996;103:1670–9.
- Asai T, Katsumori N, Mizokami K. Retinal ganglion cell damage in human glaucoma. 2. Studies on damage pattern. Nippon Ganka Gakkai Zasshi. 1987;91:1204–13.
- Garcia-Valenzuela E, Shareef S, Walsh J, Sharma SC. Programmed cell death of retinal ganglion cells during experimental glaucoma. Exp Eye Res. 1995;61:33–44.
- Quigley HA, Nickells RW, Kerrigan LA, et al. Retinal ganglion cell death in experimental glaucoma and after axotomy occurs by apoptosis. Invest Ophthalmol Vis Sci. 1995;36:774–86.

- Weber AJ, Kaufman PL, Hubbard WC. Morphology of single ganglion cells in the glaucomatous primate retina. Invest Ophthalmol Vis Sci. 1998;39: 2304–20.
- Quigley HA, McKinnon SJ, Zack DJ, et al. Retrograde axonal transport of BDNF in retinal ganglion cells is blocked by acute IOP elevation in rats. Invest Ophthalmol Vis Sci. 2000;41:3460–6.
- Quigley HA. Ganglion cell death in glaucoma: pathology recapitulates ontogeny. Aust N Z J Ophthalmol. 1995;23:85–91.
- Wygnanski T, Desatnik H, Quigley HA, Glovinsky Y. Comparison of ganglion cell loss and cone loss in experimental glaucoma. Am J Ophthalmol. 1995; 120:184–9.
- Panda S, Jonas JB. Decreased photoreceptor count in human eyes with secondary angle-closure glaucoma. Invest Ophthalmol Vis Sci. 1992;33:2532–6.
- Kendell KR, Quigley HA, Kerrigan LA, Pease ME, Quigley EN. Primary open-angle glaucoma is not associated with photoreceptor loss. Invest Ophthalmol Vis Sci. 1995;36:200–5.
- Nork TM, Ver Hoeve JN, Poulsen GL, et al. Swelling and loss of photoreceptors in chronic human and experimental glaucomas. Arch Ophthalmol. 2000; 118:235–45.
- Janssen P, Naskar R, Moore S, Thanos S, Thiel HJ. Evidence for glaucoma-induced horizontal cell alterations in the human retina. Ger J Ophthalmol. 1996;5:378–85.
- Yucel YH, Zhang Q, Gupta N, Kaufman PL, Weinreb RN. Loss of neurons in magnocellular and parvocellular layers of the lateral geniculate nucleus in glaucoma. Arch Ophthalmol. 2000;118:378–84.
- Yucel YH, Zhang Q, Weinreb RN, Kaufman PL, Gupta N. Atrophy of relay neurons in magno- and parvocellular layers in the lateral geniculate nucleus in experimental glaucoma. Invest Ophthalmol Vis Sci. 2001;42:3216–22.
- Yucel YH, Zhang Q, Weinreb RN, Kaufman PL, Gupta N. Effects of retinal ganglion cell loss on magno-, parvo-, koniocellular pathways in the lateral geniculate nucleus and visual cortex in glaucoma. Prog Retin Eye Res. 2003;22:465–81.
- Gaasterland D, Tanishima T, Kuwabara T. Axoplasmic flow during chronic experimental glaucoma. 1. Light and electron microscopic studies of the monkey optic nervehead during development of glaucomatous cupping. Invest Ophthalmol Vis Sci. 1978;17:838–46.
- Minckler DS, Bunt AH, Johanson GW. Orthograde and retrograde axoplasmic transport during acute ocular hypertension in the monkey. Invest Ophthalmol Vis Sci. 1977;16:426–41.
- Quigley HA, Addicks EM, Green WR, Maumenee AE. Optic nerve damage in human glaucoma. II. The site of injury and susceptibility to damage. Arch Ophthalmol. 1981;99:635–49.

- Quigley HA, Green WR. The histology of human glaucoma cupping and optic nerve damage: clinicopathologic correlation in 21 eyes. Ophthalmology. 1979;86:1803–30.
- Bellezza AJ, Rintalan CJ, Thompson HW, et al. Deformation of the lamina cribrosa and anterior scleral canal wall in early experimental glaucoma. Invest Ophthalmol Vis Sci. 2003;44:623–37.
- Burgoyne CF, Downs JC, Bellezza AJ, Hart RT. Three-dimensional reconstruction of normal and early glaucoma monkey optic nerve head connective tissues. Invest Ophthalmol Vis Sci. 2004; 45:4388–99.
- Downs JC, Suh JK, Thomas KA, et al. Viscoelastic material properties of the peripapillary sclera in normal and early-glaucoma monkey eyes. Invest Ophthalmol Vis Sci. 2005;46:540–6.
- Downs JC, Yang H, Girkin C, et al. Three dimensional histomorphometry of the normal and early glaucomatous monkey optic nerve head: neural canal and subarachnoid space architecture. Invest Ophthalmol Vis Sci. 2007;48:3195–208.
- 29. Yang H, Downs JC, Girkin C, et al. 3-D histomorphometry of the normal and early glaucomatous monkey optic nerve head: lamina cribrosa and peripapillary scleral position and thickness. Invest Ophthalmol Vis Sci. 2007;48:4597–607.
- 30. Yang H, Downs JC, Bellezza AJ, Thompson H, Burgoyne CF. 3-D histomorphometry of the normal and early glaucomatous monkey optic nerve head: prelaminar neural tissues and cupping. Invest Ophthalmol Vis Sci. 2007;48:5068–84.
- Johnson EC, Morrison JC, Farrell S, et al. The effect of chronically elevated intraocular pressure on the rat optic nerve head extracellular matrix. Exp Eye Res. 1996;62:663–74.
- 32. Johnson EC, Deppmeier LM, Wentzien SK, Hsu I, Morrison JC. Chronology of optic nerve head and retinal responses to elevated intraocular pressure. Invest Ophthalmol Vis Sci. 2000;41:431–42.
- Cepurna WO, Kayton RJ, Johnson EC, Morrison JC. Age related optic nerve axonal loss in adult brown Norway rats. Exp Eye Res. 2005;80:877–84.
- Howell GR, Libby RT, Jakobs TC, et al. Axons of retinal ganglion cells are insulted in the optic nerve early in DBA/2J glaucoma. J Cell Biol. 2007; 179:1523–37.
- Anderson DR. Ultrastructure of human and monkey lamina cribrosa and optic nerve head. Arch Ophthalmol. 1969;82:800–14.
- Morrison J, L'Hernault N, Jerdan J, Quigley H. Ultrastructural localization of extracellular matrix components in the monkey optic nerve head. Ubvest logtgaknik Vus Scu. 1988;29:353.
- Quigley HA, Dorman-Pease ME, Brown AE. Quantitative study of collagen and elastin of the optic nerve head and sclera in human and experimental monkey glaucoma. Curr Eye Res. 1991;10:877–88.

- Hernandez MR. Ultrastructural immunocytochemical analysis of elastin in the human lamina cribrosa. Changes in elastic fibers in primary open-angle glaucoma. Invest Ophthalmol Vis Sci. 1992;33: 2891–903.
- Fechtner RD, Weinreb RN. Mechanisms of optic nerve damage in primary open angle glaucoma. Surv Ophthalmol. 1994;39:23–42.
- Burgoyne CF, Morrison JC. The anatomy and pathophysiology of the optic nerve head in glaucoma. J Glaucoma. 2001;10:S16–8.
- 41. Burgoyne CF, Downs JC, Bellezza AJ, Suh JK, Hart RT. The optic nerve head as a biomechanical structure: a new paradigm for understanding the role of IOP-related stress and strain in the pathophysiology of glaucomatous optic nerve head damage. Prog Retin Eye Res. 2005;24:39–73.
- Bito LZ. Impact of intraocular pressure on venous outflow from the globe: a hypothesis regarding IOPdependent vascular damage in normal-tension and hypertensive glaucoma. J Glaucoma. 1996;5: 127–34.
- Langham M. The temporal relation between intraocular pressure and loss of vision in chronic simple glaucoma. Glaucoma. 1980;2:427–35.
- Hayreh SS. Pathogenesis of cupping of the optic disc. Br J Ophthalmol. 1974;58:863–76.
- Hayreh SS. Pathogenesis of optic nerve head changes in glaucoma. Semin Ophthalmol. 1986;1: 1–13.
- 46. Levkovitch-Verbin H, Quigley HA, Kerrigan-Baumrind LA, et al. Optic nerve transection in monkeys may result in secondary degeneration of retinal ganglion cells. Invest Ophthalmol Vis Sci. 2001;42:975–82.
- Wax MB, Tezel G, Edward PD. Clinical and ocular histopathological findings in a patient with normalpressure glaucoma. Arch Ophthalmol. 1998;116:993–1001.
- Wax MB. Is there a role for the immune system in glaucomatous optic neuropathy? Curr Opin Ophthalmol. 2000;11:145–50.
- 49. Schwartz M. Lessons for glaucoma from other neurodegenerative diseases: can one treatment suit them all? J Glaucoma. 2005;14:321–3.
- Schwartz M, Yoles E. Self-destructive and selfprotective processes in the damaged optic nerve: implications for glaucoma. Invest Ophthalmol Vis Sci. 2000;41:349–51.
- Anderson MG, Libby RT, Gould DB, Smith RS, John SW. High-dose radiation with bone marrow transfer prevents neurodegeneration in an inherited glaucoma. Proc Natl Acad Sci U S A. 2005;102: 4566–71.
- Pederson JE, Anderson DR. The mode of progressive disc cupping in ocular hypertension and glaucoma. Arch Ophthalmol. 1980;98:490–5.
- Pederson JE, Gaasterland DE. Laser-induced primate glaucoma. I. Progression of cupping. Arch Ophthalmol. 1984;102:1689–92.

- Johns KJ, Leonard-Martin T, Feman SS. The effect of panretinal photocoagulation on optic nerve cupping. Ophthalmology. 1989;96:211–6.
- 55. Klein BE, Klein R, Lee KE, Hoyer CJ. Does the intraocular pressure effect on optic disc cupping differ by age? Trans Am Ophthalmol Soc. 2006;104: 143–8.
- 56. Sponsel WE, Shoemaker J, Trigo Y, et al. Frequency of sustained glaucomatous-type visual field loss and associated optic nerve cupping in Beaver Dam, Wisconsin. Clin Experiment Ophthalmol. 2001; 29:352–8.
- Greenfield DS, Siatkowski RM, Glaser JS, Schatz NJ, Parrish 2nd RK. The cupped disc. Who needs neuroimaging? Ophthalmology. 1998;105:1866–74.
- Bianchi-Marzoli S, Rizzo 3rd JF, Brancato R, Lessell S. Quantitative analysis of optic disc cupping in compressive optic neuropathy. Ophthalmology. 1995;102:436–40.
- Schwartz JT, Reuling FH, Garrison RJ. Acquired cupping of the optic nerve head in normotensive eyes. Br J Ophthalmol. 1975;59:216–22.
- 60. Kalvin NH, Hamasaki DI, Gass JD. Experimental glaucoma in monkeys. I. Relationship between intraocular pressure and cupping of the optic disc and cavernous atrophy of the optic nerve. Arch Ophthalmol. 1966;76:82–93.
- Vrabec F. Glaucomatous cupping of the human optic disk: a neuro-histologic study. Albrecht Von Graefes Arch Klin Exp Ophthalmol. 1976;198:223–34.
- Anderson DR, Cynader MS. Glaucomatous optic nerve cupping as an optic neuropathy. Clin Neurosci. 1997;4:274–8.
- Quigley H, Anderson DR. Cupping of the optic disc in ischemic optic neuropathy. Trans Am Acad Ophthalmol Otolaryngol. 1977;83:755–62.
- Trobe JD, Glaser JS, Cassady J, Herschler J, Anderson DR. Nonglaucomatous excavation of the optic disc. Arch Ophthalmol. 1980;98:1046–50.
- Hayreh SS, Jonas JB. Optic disc morphology after arteritic anterior ischemic optic neuropathy. Ophthalmology. 2001;108:1586–94.
- 66. Hall ER, Klein BE, Knudtson MD, Meuer SM, Klein R. Age-related macular degeneration and optic disk cupping: the Beaver Dam Eye Study. Am J Ophthalmol. 2006;141:494–7.
- Piette SD, Sergott RC. Pathological optic-disc cupping. Curr Opin Ophthalmol. 2006;17:1–6.
- Alward WL. Macular degeneration and glaucomalike optic nerve head cupping. Am J Ophthalmol. 2004;138:135–6.
- 69. Danesh-Meyer HV, Savino PJ, Sergott RC. The prevalence of cupping in end-stage arteritic and nonarteritic anterior ischemic optic neuropathy. Ophthalmology. 2001;108:593–8.
- Ambati BK, Rizzo 3rd JF. Nonglaucomatous cupping of the optic disc. Int Ophthalmol Clin. 2001;41:139–49.
- Greenfield DS. Glaucomatous versus nonglaucomatous optic disc cupping: clinical differentiation. Semin Ophthalmol. 1999;14:95–108.

- Sharma M, Volpe NJ, Dreyer EB. Methanol-induced optic nerve cupping. Arch Ophthalmol. 1999;117:286.
- Manor RS. Documented optic disc cupping in compressive optic neuropathy. Ophthalmology. 1995;102:1577–8.
- 74. Orgul S, Gass A, Flammer J. Optic disc cupping in arteritic anterior ischemic optic neuropathy. Ophthalmologica. 1994;208:336–8.
- Sonty S, Schwartz B. Development of cupping and pallor in posterior ischemic optic neuropathy. Int Ophthalmol. 1983;6:213–20.
- Votruba M, Thiselton D, Bhattacharya SS. Optic disc morphology of patients with OPA1 autosomal dominant optic atrophy. Br J Ophthalmol. 2003;87:48–53.
- Quigley HA, Addicks EM. Chronic experimental glaucoma in primates. II. Effect of extended intraocular pressure elevation on optic nerve head and axonal transport. Invest Ophthalmol Vis Sci. 1980;19:137–52.
- Hernandez MR. The optic nerve head in glaucoma: role of astrocytes in tissue remodeling. Prog Retin Eye Res. 2000;19:297–321.
- 79. Agapova OA, Kaufman PL, Lucarelli MJ, Gabelt BT, Hernandez MR. Differential expression of matrix metalloproteinases in monkey eyes with experimental glaucoma or optic nerve transection. Brain Res. 2003;967:132–43.
- Johnson EC, Jia L, Cepurna WO, Doser TA, Morrison JC. Global changes in optic nerve head gene expression after exposure to elevated intraocular pressure in a rat glaucoma model. Invest Ophthalmol Vis Sci. 2007;48:3161–77.
- Jonas JB, Dichtl A. Optic disc morphology in myopic primary open-angle glaucoma. Graefes Arch Clin Exp Ophthalmol. 1997;235:627–33.
- Fernandez MC, Jonas JB, Naumann GO. Parapapillary chorioretinal atrophy in eyes with shallow glaucomatous optic disk cupping. Fortschr Ophthalmol. 1990;87:457–60.
- Burgoyne CF, Downs JC. Optic nerve head (ONH) biomechanics underlies the clinical behaviour and susceptibility of the aged optic nerve head. J Glaucoma. 2008;17:318–28.
- Rochtchina E, Mitchell P, Wang JJ. Relationship between age and intraocular pressure: the Blue Mountains Eye Study. Clin Experiment Ophthalmol. 2002;30:173–5.
- Nomura H, Ando F, Niino N, Shimokata H, Miyake Y. The relationship between age and intraocular pressure in a Japanese population: the influence of central corneal thickness. Curr Eye Res. 2002; 24:81–5.
- Nomura H, Shimokata H, Ando F, Miyake Y, Kuzuya F. Age-related changes in intraocular pressure in a large Japanese population: a cross-sectional and longitudinal study. Ophthalmology. 1999;106:2016–22.
- Klein BE, Klein R, Linton KL. Intraocular pressure in an American community. The Beaver Dam Eye Study. Invest Ophthalmol Vis Sci. 1992;33:2224–8.

- Weih LM, Mukesh BN, McCarty CA, Taylor HR. Association of demographic, familial, medical, and ocular factors with intraocular pressure. Arch Ophthalmol. 2001;119:875–80.
- Leske MC, Connell AM, Wu SY, Hyman L, Schachat AP. Distribution of intraocular pressure. The Barbados Eye Study. Arch Ophthalmol. 1997;115: 1051–7.
- Wu SY, Leske MC. Associations with intraocular pressure in the Barbados Eye Study. Arch Ophthalmol. 1997;115:1572–6.
- Tielsch JM, Sommer A, Katz J, et al. Racial variations in the prevalence of primary open-angle glaucoma. The Baltimore Eye Survey. J Am Med Assoc. 1991;266:369–74.
- 92. Suzuki Y, Iwase A, Araie M, et al. Risk factors for open-angle glaucoma in a Japanese population: the Tajimi Study. Ophthalmology. 2006;113: 1613–7.
- 93. Friedman DS, Jampel HD, Munoz B, West SK. The prevalence of open-angle glaucoma among blacks and whites 73 years and older: the Salisbury Eye Evaluation Glaucoma Study. Arch Ophthalmol. 2006;124:1625–30.
- Chumbley LC, Brubaker RF. Low-tension glaucoma. Am J Ophthalmol. 1976;81:761–7.
- Drance SM, Sweeney VP, Morgan RW, Feldman F. Studies of factors involved in the production of low tension glaucoma. Arch Ophthalmol. 1973;89:457–65.
- Goldberg I, Hollows FC, Kass MA, Becker B. Systemic factors in patients with low-tension glaucoma. Br J Ophthalmol. 1981;65:56–62.
- Klein BE, Klein R, Sponsel WE, et al. Prevalence of glaucoma. The Beaver Dam Eye Study. Ophthalmology. 1992;99:1499–504.
- Levene RZ. Low tension glaucoma: a critical review and new material. Surv Ophthalmol. 1980;24: 621–64.
- Shiose Y. Prevalence and clinical aspects of lowtension glaucoma. Philadelphia: Lippincott; 1983.
- Geijssen HC. Studies on normal pressure glaucoma. Amstelveen: Kugler; 1991.
- 101. Gordon MO, Beiser JA, Brandt JD, et al. The Ocular Hypertension Treatment Study: baseline factors that predict the onset of primary open-angle glaucoma. Arch Ophthalmol. 2002;120:714–20; discussion 829–730.
- 102. Nouri-Mahdavi K, Hoffman D, Coleman AL, et al. Predictive factors for glaucomatous visual field progression in the Advanced Glaucoma Intervention Study. Ophthalmology. 2004;111:1627–35.
- 103. Heijl A, Leske MC, Bengtsson B, Hussein M. Measuring visual field progression in the Early Manifest Glaucoma Trial. Acta Ophthalmol Scand. 2003;81:286–93.
- 104. Greene PR. Mechanical considerations in myopia: relative effects of accommodation, convergence, intraocular pressure, and the extraocular muscles. Am J Optom Physiol Opt. 1980;57:902–14.

- 105. Bellezza AJ, Hart RT, Burgoyne CF. The optic nerve head as a biomechanical structure: initial finite element modeling. Invest Ophthalmol Vis Sci. 2000;41:2991–3000.
- 106. Sigal IA, Flanagan JG, Ethier CR. Factors influencing optic nerve head biomechanics. Invest Ophthalmol Vis Sci. 2005;46:4189–99.
- 107. Sigal IA, Flanagan JG, Tertinegg I, Ethier CR. Reconstruction of human optic nerve heads for finite element modeling. Technol Health Care. 2005;13:313–29.
- Sigal IA, Flanagan JG, Tertinegg I, Ethier CR. Finite element modeling of optic nerve head biomechanics. Invest Ophthalmol Vis Sci. 2004;45:4378–87.
- Repka MX, Quigley HA. The effect of age on normal human optic nerve fiber number and diameter. Ophthalmology. 1989;96:26–32.
- 110. Balazsi AG, Rootman J, Drance SM, Schulzer M, Douglas GR. The effect of age on the nerve fiber population of the human optic nerve. Am J Ophthalmol. 1984;97:760–6.
- 111. Morrison JC, Cork LC, Dunkelberger GR, Brown A, Quigley HA. Aging changes of the rhesus monkey optic nerve. Invest Ophthalmol Vis Sci. 1990; 31:1623–7.
- 112. Cull G, Cioffi GA, Dong J, Homer L, Wang L. Estimating normal optic nerve axon numbers in non-human primate eyes. J Glaucoma. 2003;12: 301–6.
- Sandell JH, Peters A. Effects of age on the glial cells in the rhesus monkey optic nerve. J Comp Neurol. 2002;445:13–28.
- 114. Sandell JH, Peters A. Effects of age on nerve fibers in the rhesus monkey optic nerve. J Comp Neurol. 2001;429:541–53.
- 115. Frisen L. High-pass resolution perimetry and agerelated loss of visual pathway neurons. Acta Ophthalmol (Copenh). 1991;69:511–5.
- Albon J, Purslow PP, Karwatowski WS, Easty DL. Age related compliance of the lamina cribrosa in human eyes. Br J Ophthalmol. 2000;84:318–23.
- 117. Morrison JC, Jerdan JA, Dorman ME, Quigley HA. Structural proteins of the neonatal and adult lamina cribrosa. Arch Ophthalmol. 1989;107: 1220–4.
- Pena JD, Roy S, Hernandez MR. Tropoelastin gene expression in optic nerve heads of normal and glaucomatous subjects. Matrix Biol. 1996;15:323–30.
- Quigley HA. Childhood glaucoma: results with trabeculotomy and study of reversible cupping. Ophthalmology. 1982;89:219–26.
- 120. Hernandez MR, Luo XX, Andrzejewska W, Neufeld AH. Age-related changes in the extracellular matrix of the human optic nerve head. Am J Ophthalmol. 1989;107:476–84.
- 121. Jeffery G, Evans A, Albon J, et al. The human optic nerve: fascicular organisation and connective tissue types along the extra-fascicular matrix. Anat Embryol (Berl). 1995;191:491–502.

- 122. Albon J, Karwatowski WS, Easty DL, Sims TJ, Duance VC. Age related changes in the noncollagenous components of the extracellular matrix of the human lamina cribrosa. Br J Ophthalmol. 2000;84:311–7.
- Bailey AJ, Paul RG, Knott L. Mechanisms of maturation and ageing of collagen. Mech Ageing Dev. 1998;106:1–56.
- 124. Brown CT, Vural M, Johnson M, Trinkaus-Randall V. Age-related changes of scleral hydration and sulfated glycosaminoglycans. Mech Ageing Dev. 1994;77:97–107.
- 125. Albon J, Karwatowski WS, Avery N, Easty DL, Duance VC. Changes in the collagenous matrix of the aging human lamina cribrosa. Br J Ophthalmol. 1995;79:368–75.
- 126. Friedenwald J. It is evident that an increasing rigidity of the ocular coats is characteristic of advancing age. AJO. 1937;20:985–1024.
- 127. Kotecha A, Izadi S, Jeffrey G. Age related changes in the thickness of the human lamina cribrosa. Br J Ophthalmol. 2006;90:1531–4.
- Albon J, Farrant S, Akhtar S, et al. Connective tissue structure of the tree shrew optic nerve and associated ageing changes. Invest Ophthalmol Vis Sci. 2007;48:2134–44.
- 129. Grunwald JE, Piltz J, Patel N, Bose S, Riva CE. Effect of aging on retinal macular microcirculation: a blue field simulation study. Invest Ophthalmol Vis Sci. 1993;34:3609–13.
- 130. Harris A, Harris M, Biller J, et al. Aging affects the retrobulbar circulation differently in women and men. Arch Ophthalmol. 2000;118:1076–80.
- 131. Hernandez MR, Wang N, Hanley NM, Neufeld AH. Localization of collagen types I and IV mRNAs in human optic nerve head by in situ hybridization. Invest Ophthalmol Vis Sci. 1991;32: 2169–77.
- 132. May CA. The optic nerve head region of the aged rat: an immunohistochemical investigation. Curr Eye Res. 2003;26:347–54.
- 133. Investigators TA. The advanced glaucoma intervention study (AGIS): 7. The relationship between control of intraocular pressure and visual field deterioration. Am J Ophthalmol. 2000;130:429–40.
- 134. Kass MA, Heuer DK, Higginbotham EJM, et al. The ocular hypertension treatment study (a randomized trial determines that topical ocular hypotensive medication delays or prevents the onset of primary openangle glaucoma). Arch Ophthalmol. 2002;120: 701–13.
- 135. Leske MC, Heijl A, Hussein M, et al. Factors for glaucoma progression and the effect of treatment: the early manifest glaucoma trial. Arch Ophthalmol. 2003;121:48–56.
- 136. Anderson DR, Drance SM, Schulzer M. Factors that predict the benefit of lowering intraocular pressure in normal tension glaucoma. Am J Ophthalmol. 2003;136:820–9.

- 137. Zeimer R. Could glaucoma damage be due to a viscoelastic mismatch between the sclera and the lamina cribrosa? J Jpn Glaucoma Soc. 1992;2:17–20.
- 138. Zeimer R. Biomechanical properties of the optic nerve head. In: Drance SM, editor. Optic nerve in glaucoma. Amsterdam: Kugler; 1995. p. 107–21.
- 139. Jonas JB, Berenshtein E, Holbach L. Lamina cribrosa thickness and spatial relationships between intraocular space and cerebrospinal fluid space in highly myopic eyes. Invest Ophthalmol Vis Sci. 2004;45:2660–5.
- 140. Cioffi GA, Van Buskirk EM. Vasculature of the anterior optic nerve and peripapillary choroid. 2nd ed. St. Louis: Mosby; 1996. p. 177–97.
- 141. Quigley HA. Overview and introduction to session on connective tissue of the optic nerve in glaucoma.

Chapter 2. In: Drance SM, Anderson DR, editors. Optic nerve in glaucoma. Amsterdam: Kugler Publications; 1995. p. 15–36.

- 142. Morrision JC, L'Hemault NL, Jerdan JA, Quigley HA. Ultrastructural location of extracellular matrix components in the optic nerve head. Arch Opthamol. 1989;107:123–9.
- 143. Emery JM, Landis D, Paton D, Boniuk M, Craig JM. The lamina cribrosa in normal and glaucomatous human eyes. Trans Am Acad Ophthalmol Otolaryngol. 1974;78:OP290–7.
- 144. Quigley HA, Hohman RM, Addicks EM, Massof RW, Green WR. Morphologic changes in the lamina cribrosa correlated with neural loss in open-angle glaucoma. Am J Ophthalmol. 1983;95:673–91.